REVIEW

Endocannabinoids and cannabinoid receptors in ischaemia-reperfusion injury and preconditioning

P Pacher¹ and G Haskó²

¹Section on Oxidative Stress Tissue Injury, Laboratory of Physiological Studies, NIAAA, National Institutes of Health, Bethesda, MD, USA and ²Department of Surgery, UMDNJ-New Jersey Medical School, Newark, NJ, USA

Ischaemia-reperfusion (I/R) is a pivotal mechanism of organ injury during stroke, myocardial infarction, organ transplantation and vascular surgeries. Ischaemic preconditioning (IPC) is a potent endogenous form of tissue protection against I/R injury. On the one hand, endocannabinoids have been implicated in the protective effects of IPC through cannabinoid CB₁/CB₂ receptordependent and -independent mechanisms. However, there is evidence suggesting that endocannabinoids are overproduced during various forms of I/R, such as myocardial infarction or whole body I/R associated with circulatory shock, and may contribute to the cardiovascular depressive state associated with these pathologies. Previous studies using synthetic CB₁ receptor agonists or knockout mice demonstrated CB₁ receptor-dependent protection against cerebral I/R injury in various animal models. In contrast, several follow-up reports have shown protection afforded by CB₁ receptor antagonists, but not agonists. Excitedly, emerging studies using potent CB2 receptor agonists and/or knockout mice have provided compelling evidence that CB₂ receptor activation is protective against myocardial, cerebral and hepatic I/R injuries by decreasing the endothelial cell activation/inflammatory response (for example, expression of adhesion molecules, secretion of chemokines, and so on), and by attenuating the leukocyte chemotaxis, rolling, adhesion to endothelium, activation and transendothelial migration, and interrelated oxidative/nitrosative damage. This review is aimed to discuss the role of endocannabinoids and CB receptors in various forms of I/R injury (myocardial, cerebral, hepatic and circulatory shock) and preconditioning, and to delineate the evidence supporting the therapeutic utility of selective CB₂ receptor agonists, which are devoid of psychoactive effects, as a promising new approach to limit I/R-induced tissue damage.

British Journal of Pharmacology (2008) 153, 252-262; doi:10.1038/sj.bjp.0707582; published online 19 November 2007

Keywords: ischaemia-reperfusion; preconditioning; endocannabinoids; cannabinoid receptors; inflammation; leukocyte chemotaxis and adhesion

Abbreviations: AEA, anandamide; 2-AG, 2-arachidonoylqlycerol; CB, cannabinoid; IPC, ischaemic preconditioning; I/R, ischaemia-reperfusion

Introduction

Ischaemic-reperfusion (I/R) injury is the principal cause of tissue damage occurring in conditions such as stroke, myocardial infarction, cardiopulmonary bypass and other vascular surgeries, and organ transplantation, as well as a major mechanism of end-organ damage complicating the course of circulatory shock of various aetiologies. In all these conditions, the initial trigger of the damage is the transient disruption of the normal blood supply to target organs followed by reperfusion. Reperfusion of ischaemic tissues is the ultimate treatment to reduce tissue injury. Unfortunately, reperfusion itself inflicts additional tissue damage mediated by reactive oxygen (superoxide anion, hydrogen peroxide and hydroxyl radical) and reactive nitrogen species (for example, peroxynitrite) upon reperfusion, as well as to the rapid transcriptional activation of an array of proinflammatory genes (reviewed in Ferdinandy and Schulz, 2003; Podgoreanu et al., 2005; Pacher et al., 2005e, 2007; Ungvari et al., 2005). Direct consequences are the local sequestration and activation of polymorphonuclear leukocytes, leading to a rapid amplification of the initial inflammatory response and reactive oxygen species generation, so-called 'respiratory burst' (Lucchesi, 1990). Additional sources of increased reactive oxygen species generation during I/R can be xanthine and NAD(P)H oxidases, mitochondria, COX and uncoupled nitric oxide synthases (reviewed in Griendling et al., 2000; Ungvari et al., 2005; Pacher et al., 2006b). The burst of reactive oxygen and nitrogen species immediately upon reperfusion initiates a chain of deleterious cellular responses eventually leading to endothelial inflammatory response and dysfunction, adherence of neutrophils and

Correspondence: Dr P Pacher, Section on Oxidative Stress and Tissue Injury, Laboratory of Physiological Studies, NIAAA, National Institutes of Health, 5625 Fishers Lane, MSC-9413, Bethesda, MD 20892-9413, USA.

E-mail: pacher@mail.nih.gov

Received 16 July 2007; revised 28 August 2007; accepted 30 August 2007; published online 19 November 2007

lymphocytes to the endothelium, transendothelial migration of inflammatory cells, the release of various harmful mediators, cellular calcium overload, and eventually cell death and organ dysfunction.

Ischaemic preconditioning (IPC), first introduced by Murry et al. (1986) is a potent endogenous form of protection against I/R injury. In hearts and various other organs, IPC (brief episode(s) of ischaemia applied before the main I/R) reduces infarct size and enhances the recovery of organ function (Yellon and Hausenloy, 2005). Preconditioning can also be achieved with bacterial endotoxins and various other chemicals and when brief episodes of ischaemia are applied following the ischaemic period (the latter is called postconditioning (for reviews see Yellon and Hausenloy, 2005; Bolli, 2007; Hausenloy and Yellon, 2007).

Circulatory shock classifies a syndrome precipitated by a systemic derangement in perfusion leading to widespread cellular hypoxia and vital organ dysfunction. Dependent on its initial pathophysiological mechanisms, shock is subdivided in three main categories, namely cardiogenic, haemorrhagic and septic shock. In the advanced stages, all shock states evolve to a common clinical picture characterized by profound tissue ischaemia, cardiovascular failure, the activation of cellular cytotoxic effectors (polymophonuclear leukocytes) and the upregulation of an array of proinflammatory genes, leading to systemic inflammation, organ dysfunction and death (Hotchkiss and Karl, 2003). From a clinical point of view, at present no therapy is available to limit reperfusion injury, which highlights the importance of

a better understanding of its underlying mechanisms, to devise better future therapeutic approaches.

Numerous studies have suggested that the endocannabinoid system may modulate I/R injury (reviewed in Lamontagne et al., 2006; Pacher et al., 2006a). Endocannabinoids have been implicated in the protective effects of IPC through cannabinoid (CB) receptor-dependent and -independent mechanisms; however, they may also contribute to the cardiovascular collapse associated with myocardial infarction and circulatory shock (reviewed in Pacher et al., 2006a). To date, two CB receptors have been identified by molecular cloning: the CB₁ and CB₂ receptors. The CB₁ receptor is abundantly expressed in brain tissue (Matsuda et al., 1990), but is also present in peripheral tissues including vasculature (Gebremedhin et al., 1999; Liu et al., 2000), heart (Batkai et al., 2004b; Pacher et al., 2005d) and liver (Batkai et al., 2001; Engeli et al., 2005; Osei-Hyiaman et al., 2005; Teixeira-Clerc et al., 2006). The CB2 receptor was previously considered to be expressed primarily in immune and haematopoietic cells (Munro et al., 1993; reviewed in Pacher et al., 2006a). However, more recent studies have also found CB₂ receptors in brain (Van Sickle et al., 2005), myocardium (Mukhopadhyay et al., 2007), cardiomyoblasts (Shmist et al., 2006; Mukhopadhyay et al., 2007) and endothelial cells of various origins (Blazquez et al., 2003; Zoratti et al., 2003; Golech et al., 2004; Mestre et al., 2006; Rajesh et al., 2007a, b). The synthetic and natural ligands (the latter called endocannabinoids: arachidonoyl ethanolamide or anandamide (AEA) and 2-arachidonoylglycerol (2-AG)) of CB

Table 1 Ranges of K_i values for certain cannabinoid CB_1 and/or CB_2 receptor agonists or antagonists/inverse agonists for the *in vitro* displacement of [3 H]CP55940, [3 H]HU243 or [3 H]BAY38-7271 from CB_1 - and CB_2 -specific binding sites (based on Howlett *et al.*, 2002; Pertwee, 2005a; for details see therein)

| Agonist/ligand | CB_1 K _i value (nM) | CB_2 K_i value (nM) | Reference |
|--|----------------------------------|-------------------------|-----------------------------|
| CB ₁ -selective agonists | | | |
| ACEA | 1.4–5.29 | 195 to > 2000 | Pertwee (2005a) |
| R-(+)-methanandamide | 17.9–28.3 | 815–868 | Pertwee (2005a) |
| Agonists without any marked CB ₁ /CB ₂ | selectivity | | |
| Anandamide | 61–543 | 279–1940 | Pertwee (2005a) |
| BAY38-7271 | 1.85 | 5.96 | Pertwee (2005a) |
| 2-Arachidonoyl glycerol | 58.3-472 | 145–1400 | Pertwee (2005a) |
| HU-210 | 0.0608-0.1 | 0.17-0.524 | Pertwee (2005a) |
| CP55940 | 0.5–5 | 0.69-2.8 | Pertwee (2005a) |
| Δ^9 -THC | 5.05-53.3 | 3.13–75.3 | Pertwee (2005a) |
| R-(+)-WIN 55,212-2 | 1.89–123 | 0.28–16.2 | Pertwee (2005a) |
| CB ₂ -selective agonists | | | |
| JWH015 | 383 | 13.8 | Pertwee (2005a) |
| JWH133 | 677 | 3.4 | Pertwee (2005a) |
| HU-308 | >10000 | 22.7 | Pertwee (2005a) |
| O-1966 | 5055 ± 984 | 23 ± 2.1 | Wiley et al. (2002) |
| O-3853 | 1509 ± 148 | 6.0 ± 2.5 | Zhang <i>et al</i> . (2007) |
| CB ₁ -selective antagonist/inverse agonis | ts | | |
| SR141716A | 1.8–11.8 | 515–13 200 | Pertwee (2005a) |
| AM281 | 12 | 4200 | Pertwee (2005a) |
| AM251 | 7.49 | 2290 | Pertwee (2005a) |
| LY320135 | 141 | 14 900 | Pertwee (2005a) |
| CB ₂ -selective antagonist/inverse agonis | ts | | |
| AM630 | 5152 | 31.2 | Pertwee (2005a) |
| SR144528 | 50.3 to > 10000 | 0.28-5.6 | Pertwee (2005a) |

P Pacher and G Haskó

receptors exert various anti-inflammatory and neuroprotective (Panikashvili et al., 2001, 2005, 2006) effects by inhibiting the generation and release of proinflammatory cytokines and mediators (reviewed in Mechoulam et al., 2002a, b; Klein, 2005; Pacher et al., 2006a). The pharmacological modulation of the endocannabinoid system represents a promising strategy in various cardiovascular, inflammatory, metabolic, gastrointestinal and liver disorders (reviewed in Di Marzo et al., 2004; Pacher et al., 2005a, 2006a; Pertwee, 2005b; Mackie, 2006; Mallat et al., 2007). The selectivity of the endocannabinoids and synthetic ligands used in various I/R studies towards CB₁/CB₂ receptors are summarized in Table 1 (for excellent detailed overviews on the subject and also on the development of CB₁/CB₂ receptor knockout mice, see Howlett et al., 2002; Pertwee, 2005a).

In this review, we will discuss the triggers and sources of endocannabinoid production during various forms of I/R injury (myocardial, cerebral, hepatic and retinal ischaemia, and circulatory shock) and preconditioning, as well as the diverse role of these novel mediators and their receptors in these processes. We will also overview the accumulating evidence obtained through the use of various synthetic CB₁/ CB₂ receptor ligands, with particular focus on the novel role of CB₂ receptors, suggesting that the modulation of the endocannabinoid system can be therapeutically exploited in various forms of I/R injury.

Myocardial I/R and preconditioning

Initial studies used isolated heart preparations to study the role of endocannabinoid system in myocardial I/R and preconditioning. Lagneux and Lamontagne (2001) implicated for the first time the involvement of the endocannabinoid system in endotoxin (lipopolysaccharide (LPS))-induced preconditioning against myocardial I/R injury, based on the assumption that LPS increases endocannabinoid production in inflammatory cells (Varga et al., 1998; Maccarrone et al., 2001; Liu et al., 2003). They compared the effects of 90 min of low-flow ischaemia followed by 60-min reperfusion at normal flow in isolated hearts from rats pretreated with LPS or saline. LPS pretreatment reduced infarct size and enhanced functional recovery upon reperfusion compared to controls, which could be attenuated by the CB₂ antagonist SR144528, but not by the CB₁ antagonist SR141716, suggesting the involvement of myocardial CB2 receptors in the observed LPS-induced cardioprotection (Lagneux and Lamontagne, 2001). In a consequent study, in which the preconditioning was triggered by heat stress, SR144528 but not SR141716 also abolished the infarct-size-reducing effect of heat stress (Joyeux et al., 2002). The conclusion of these early studies was that the protection afforded by LPS- or heat stress-induced preconditioning was mediated by endocannabinoids acting on CB2 receptors. In contrast, in preconditioning induced by a brief period of ischaemia (5 min), either CB₂ or CB₁ receptor blockade could abolish the protection, and both CB₁ and CB₂ receptors were implicated in the preservation of the endothelium-dependent, 5-HT-induced vasodilation by IPC (Bouchard et al., 2003). Palmitoylethanolamide or 2-AG, but not AEA, added to the perfusion medium of isolated rat hearts afforded protection against ischaemia by decreasing myocardial damage and infarct size and by improving myocardial functional recovery (Lepicier et al., 2003). SR144528 completely blocked the cardioprotective effect of both palmitoylethanolamide and 2-AG, whereas SR141716 only partially inhibited the effect of 2-AG only (Lepicier et al., 2003). Similarly, CB₁ and CB₂ agonists ACEA and JWH015 also reduced infarct size in this model, and the CB2 receptor-mediated cardioprotection by palmitoylethanolamide involved activation of p38/ERK (extracellular signal-regulated kinase)1/2 kinases and PKC (Lepicier et al., 2003). On the contrary, Underdown et al. (2005) have found that the infarct-size-reducing effect of AEA could be equally well antagonized by both CB₁ and CB₂ antagonists; however, it could not be mimicked by selective CB₁ or CB₂ agonists, suggesting the involvement of a site distinct from CB₁ or CB₂ receptors. Another recent study using a model of delayed preconditioning in rats induced by administration of the nitric oxide donor nitroglycerin for 24 h via transdermal application suggested that the protective effect of nitroglycerin against myocardial infarction is mediated via CB₁ receptors. Nitroglycerin increased the myocardial content of 2-AG, but not AEA (Wagner et al., 2006). The major limitation of the above-mentioned studies is the use of ex vivo models (for example, buffer-perfused isolated heart preparations) that could not address the question of whether endocannabinoids or synthetic agonists can modulate endothelial or immune cell activation and interactions, which are pivotal events in the sequel of reperfusion damage (reviewed in Pertwee, 2005a; Lamontagne et al., 2006; Pacher et al., 2006a). Despite the above-mentioned limitation, these pioneering studies importantly implied the possible contribution of functional CB2 receptors in cardiomyocytes and/or endothelial cells responsible, at least in part, to the protective effects of preconditioning. Indeed, consequent studies have demonstrated the presence of CB₂ receptors in myocardium (Mukhopadhyay et al., 2007), cardiomyoblasts (Shmist et al., 2006; Mukhopadhyay et al., 2007) and endothelial cells of various origins (Blazquez et al., 2003; Zoratti et al., 2003; Golech et al., 2004; Mestre et al., 2006; Rajesh et al., 2007a, b). Consistently with the beneficial effect of CB₂ receptor activation on cardiomyocytes, a recent study demonstrated that delta(9)-tetrahydrocannabinol (THC) protected H9c2 cardiomyoblasts subjected to hypoxia in vitro presumably via CB₂ receptor activation and increased nitric oxide production (Shmist et al., 2006).

In a clinically more relevant rat model of I/R injury, both AEA and HU-210 decreased the incidence of ventricular arrhythmias and reduced infarct size, presumably through the activation of CB₂ but not CB₁ receptors (Krylatov *et al.*, 2001, 2002a, b; Ugdyzhekova *et al.*, 2001, 2002). In a mouse model of myocardial I/R induced by coronary artery ligation, the reduction of leukocyte-dependent second wave of myocardial damage subsequent to the initial IR injury was attributed to CB₂ receptor activation, since the protection afforded by WIN 55,212-2 could be prevented by AM630, but not by the CB₁ antagonist AM251 (Di Filippo *et al.*, 2004). Two recent studies using rat models of acute and chronic

myocardial infarction demonstrated that endocannabinoids contribute to the hypotension and cardiodepression associated with acute cardiogenic shock, which could be attenuated by CB₁ antagonists (Wagner *et al.*, 2001, 2003).

Collectively, although the role of CB receptors and endocannabinoids in protection afforded by preconditioning against myocardial I/R is still a very controversial issue requiring further clarification by using knockout mice and more selective ligands for CB_2 receptors, the findings implicating the importance of CB_2 receptor, presumably both on endothelial and inflammatory cells, and perhaps on cardiomyocytes, are very encouraging.

Cerebral I/R (stroke)

Ischaemic stroke, resulting from the reduction of cerebral blood flow in the territory of a major cerebral artery due to its transient or permanent occlusion by local thrombosis or embolus is the second leading cause of death in industrialized countries and the leading medical cause of acquired adult disability. One in six patients die in the first 4 weeks following ischaemic stroke, and half of the survivors are permanently disabled in spite of the best efforts to rehabilitate them to avoid complications (Klijn and Hankey, 2003). A cascade of complex molecular events is set in motion during cerebral ischaemia and culminates in neuronal cell death. Improving our understanding of these events might help to devise novel therapies to limit neuronal injury in stroke patients, a concept termed 'neuroprotection' (Lees et al., 2006).

The endocannabinoid system may represent a pivotal neuroprotective mechanism both in acute forms of neuronal injury (for example, stroke and traumatic brain injury) and in various chronic neurodegenerative disorders, including multiple sclerosis, Parkinson's disease, Huntington's disease, Alzheimer's disease and amyotrophic lateral sclerosis (reviewed in Mechoulam et al., 2002b; Croxford, 2003; Sarne and Mechoulam, 2005; Mackie, 2006; Pacher et al., 2006a). Even though the exact mechanisms of these neuroprotective effects are not completely understood, numerous CB receptor-dependent as well as receptor-independent processes have been suggested to be involved, which include, but are not limited to: (1) modulation of immune responses and the release of inflammatory mediators by CB₁, CB₂ and non-CB₁/ CB₂ receptors on neurons, astrocytes, microglia, macrophages, neutrophils and lymphocytes (Walter and Stella, 2004; Klein, 2005); (2) modulation of synaptic plasticity and excitatory glutamatergic transmissions via presynaptic CB₁ receptors (Freund et al., 2003; Piomelli, 2003); (3) activation of cytoprotective signalling pathways (for example, PKB/Akt, PKA or neurotrophic factors) (Pacher et al., 2006a); (4) modulation of calcium homoeostasis and excitability via interactions with Ca2+, K+ and Na+ channels, gap junctions and intracellular Ca^{2+} stores, NMDA receptors (Freund et al., 2003; Piomelli, 2003; Pacher et al., 2006a); (5) CB₁ receptor-mediated central hypothermia, presumably by decreasing metabolic rate and oxygen demand; (6) antioxidant properties of CBs (Hampson et al., 2000); (7) modulation of endothelial activation and inflammatory response, leukocyte rolling, adhesion to the endothelium, transmigration and activation presumably by CB₂ receptors.

The first evidence for the neuroprotective effect of CBs came from the stroke research field from studies using synthetic non-psychotropic CB Dexanabinol/HU-211, which exerted its beneficial effects through CB₁/CB₂-independent mechanisms, in various rat and gerbil models of in vivo cerebral ischaemia (reviewed in Pacher et al., 2006a). Followup studies have also investigated the neuroprotective effects of CB₁ receptor stimulation using synthetic agonists. The synthetic CB WIN 55,212-2 attenuated hippocampal neuronal loss following transient global cerebral ischaemia in rats and reduced infarct size after permanent focal cerebral ischaemia induced by middle cerebral artery occlusion, when given 40 min prior or 30 min after the occlusion, in a CB₁-dependent manner, since the protective effect was preventable by SR141716 (Nagayama et al., 1999). WIN 55,212-2, as well as AEA and 2-AG, also protected cultured cerebral cortical neurons from in vitro glucose deprivation and hypoxia, but these effects were insensitive to CB₁ and CB₂ receptor antagonists (Nagayama et al., 1999; Sinor et al., 2000). In rat models of middle cerebral artery occlusion another synthetic agonist BAY38-7271 reduced infarct size even when given intravenously 4h following the occlusion (Mauler et al., 2002). Similarly, HU-210 improved motor disability and decreased infarct size by up to 77% in a similar model (Leker et al., 2003). Pretreatment with SR141716 partially attenuated the protective effect of HU-210 indicating CB₁ receptor involvement. However, the protective effect of HU-210 could be abolished completely by warming the animals to the body temperature of controls, indicating that the CB₁-mediated hypothermia was responsible for the observed beneficial effects (Leker et al., 2003). Similarly, CB₁mediated hypothermia was responsible for the neuroprotective effects of $\Delta(9)$ -tetrahydrocannabinol in a mouse ischaemic model of cerebral injury (Hayakawa et al., 2004) and, perhaps, also in a rat model of global cerebral ischaemia (Louw et al., 2000). Consistent with CB₁-mediated cerebroprotection, CB₁ knockout mice had increased neurotoxicity to NMDA and elevated mortality from permanent focal cerebral ischaemia, increased infarct size, more severe neurological deficits after transient focal cerebral ischaemia and decreased cerebral blood flow in the ischaemic penumbra during reperfusion, as compared to wild-type controls subjected to the same insult (Parmentier-Batteur et al., 2002).

In contrast, several more recent studies do not support the neuroprotective role of endocannabinoids and CB₁ receptor activation. In fact, the CB₁ antagonists SR141716 and LY320135 were found to reduce infarct size and to improve neurological function in a rat model of cerebral ischaemia induced by middle cerebral artery occlusion (Berger *et al.*, 2004; Muthian *et al.*, 2004; Sommer *et al.*, 2006), whereas low doses of WIN 55,212-2 had no protective effect (Muthian *et al.*, 2004).

Recent studies have also evaluated the effects of selective CB_2 agonists (O-3853, O-1966) in a stroke model. CB_2 agonists significantly decreased cerebral infarction and improved motor function after 1h middle cerebral artery occlusion followed by 23h reperfusion in mice, by attenuating the transient ischaemia-induced increase in leukocyte rolling and

adhesion to vascular endothelial cells (Zhang *et al.*, 2007). The role of CB₂ receptors in I/R injury was further supported by increased accumulation of CB₂-positive macrophages derived from resident microglia and/or invading monocytes following

cerebral I/R (Ashton et al., 2007).

Collectively, it appears that both CB_1 agonists and antagonists may afford neuroprotective effects against cerebral I/R. The reason for the contradictory effects of pharmacological blockade vs genetic knockout of CB_1 receptors is not clear, and may be related to CB_1 receptor-independent effects of antagonists, but this issue needs further clarification. In the case of CB_2 agonists, the most likely mechanism of protection is the attenuation of the transient I/R-induced increase in leukocyte infiltration, rolling and adhesion to vascular endothelial cells, and consequent activation.

Circulatory shock (full organ/body ischaemia and/or I/R)

In addition to their well-known immunological and neurobehavioral actions, CBs and their endogenous and synthetic analogues exert complex cardiodepressive and vasodilatory effects, which have been implicated in the mechanism of hypotension associated with haemorrhagic (Wagner et al., 1997; Cainazzo et al., 2002), endotoxic (Varga et al., 1998; Liu et al., 2003, 2006; Batkai et al., 2004a; Kadoi and Goto, 2006), septic (Kadoi et al., 2005), and cardiogenic shock (Wagner et al., 2001, 2003), advanced liver cirrhosis (Batkai et al., 2001; Ros et al., 2002), cirrhotic cardiomyopathy (Gaskari et al., 2005; Pacher et al., 2005c; Moezi et al., 2006; Yang et al., 2007; Batkai et al., 2007a), doxorubicin-induced heart failure (Mukhopadhyay et al., 2007) and the shock associated with necrotizing pancreatitis (Matsuda et al., 2005). Importantly, these cardiovascular depressive effects could be prevented or reversed by pretreatment with CB₁ antagonists, and are subjects of numerous comprehensive recent overviews (Randall et al., 2002; Hiley and Ford, 2004; Lamontagne et al., 2006; Lepicier et al., 2006; Mallat et al., 2007; Mendizabal and Adler-Graschinsky, 2007; Pacher et al., 2005a, b, 2006a). CB receptor antagonists (for example, SR141716, AM281, AM251 and SR144528) prolonged survival in endotoxic and septic shock or necrotizing pancreatitis (Varga et al., 1998; Smith et al., 2001; Cainazzo et al., 2002; Kadoi et al., 2005; Matsuda et al., 2005; Kadoi and Goto, 2006), while increased mortality in haemorrhagic (Wagner et al., 1997) and cardiogenic shock (Wagner et al., 2001) despite the increase in blood pressure. One explanation for this puzzling controversy is that endocannabinoidmediated vasodilation may have survival value through improving tissue oxygenation by counteracting the excessive sympathetic vasoconstriction triggered by haemorrhage or myocardial infarction, and this would be removed by CB₁ blockade. In contrast, CB₁ blockade may improve survival in endotoxic shock by preventing the primary hypotensive response to LPS (Randall et al., 2002; Hiley and Ford, 2004; Mendizabal and Adler-Graschinsky, 2007; Pacher et al., 2005a, b, 2006a). Complicating the picture, in haemorrhagic, cardiogenic and endotoxic shock, the CB agonists HU-210, WIN 55,212-2 and THC also improved endothelial function and/or survival (Wagner et al., 1997, 2001; Varga et al., 1998; Smith et al., 2000, 2001). Since the cardiovascular dysfunction and failure in most of the abovementioned conditions are triggered by overwhelming tissue ischaemia and/or I/R, and consequent oxidative/nitrosative stress and inflammatory response coupled with the activation of various downstream cell death pathways (reviewed in Evgenov and Liaudet, 2005; Ungvari et al., 2005; Pacher et al., 2005e, 2007), another explanation for the diverse beneficial effects of both agonists and antagonists in circulatory shock could lie in their various anti-inflammatory and/or antioxidant properties (reviewed in Walter and Stella, 2004; Klein, 2005), which may be attributed to their inverse agonistic properties or to CB_{1/2} receptor-independent mechanisms (reviewed in Begg et al., 2005; Pertwee, 2005a, b, 2006).

Overall, it seems that both CBs and antagonists of CB receptors may have various favorable effects in rodent shock models; however, the specificity of these effects and the relevance to human circulatory shock should be established by further studies.

Hepatic I/R

Hepatic I/R injury continues to be a fatal complication that can follow liver surgery or transplantation. It is well known that hepatic I/R injury is dependent on polymorphonuclear cell (PMN) infiltration, Kupffer cell activation and inflammatory cytokine responses (Jaeschke et al., 1996, 1997, 2006; Ohkohchi et al., 1999). Adhesion molecules mediate the initial attachment of neutrophils to the activated endothelium (Carlos and Harlan, 1994; Jaeschke, 1997). On reperfusion, tumour necrosis factor- α (TNF- α) acts as a continuous stimulator for neutrophil infiltration in the liver and it also upregulates the production of cell-type-specific leukocyte chemoattractants, known as chemokines, which have also been shown to cause upregulation of cell adhesion molecules and neutrophil activation (Jaeschke, 2006). The increased inflammatory response further aggravates oxidative stress and initiates a chain of deleterious events eventually culminating in cellular dysfunction and death.

In two recent studies, we have investigated the involvement of the endocannabinoid system in an in vivo mouse model of hepatic I/R injury using selective CB₂ agonists and CB₂ knockout mice (Batkai et al., 2007b). Activation of CB₂ receptors by JWH133 prior to the insult protected against I/R damage (measured by serum transaminases activity) by decreasing inflammatory cell infiltration, tissue and serum TNF-α, chemokines macrophage-inflammatory protein-1α $(MIP-1\alpha)$ and macrophage-inflammatory protein-2 (MIP-2) levels, tissue lipid peroxidation, and expression of adhesion molecule intercellular adhesion molecule-1 (ICAM-1). JWH133 also decreased the TNF-α-induced ICAM-1 and vascular cell adhesion molecule-1 expressions in human liver sinusoidal endothelial cells and the adhesion of human neutrophils to human liver sinusoidal endothelial cells in vitro. In agreement with the protective role of CB₂ receptor activation, CB₂^{-/-} mice developed increased I/R-induced tissue damage and proinflammatory phenotype (Batkai

P Pacher and G Haskó

et al., 2007b). In a follow-up study, we have demonstrated that the potent CB₂ receptor agonist HU-308, given prior to the induction of I/R, significantly attenuated the extent of liver damage (measured by serum alanine aminotransferase and lactate dehydrogenase), decreased serum and tissue TNF- α , MIP-1 α and MIP-2 levels, tissue lipid peroxidation, neutrophil infiltration, DNA fragmentation and caspase 3 activity. The protective effect of HU-308 against liver damage was also preserved when given immediately after the ischaemic episode. CB2 receptor was expressed in human liver sinusoidal endothelial cells and its activation by HU-308 also attenuated the TNF-α-induced ICAM-1 and vascular cell adhesion molecule-1 expression and the adhesion of human neutrophils to human liver sinusoidal endothelial cells in vitro. These findings, coupled with recent results from myocardial (Di Filippo et al., 2004) and cerebral I/R models (Zhang et al., 2007), and antifibrotic effects of CB₂ receptor in the liver (Julien et al., 2005), suggest that selective CB₂ receptor agonists may represent a novel protective strategy against hepatic and other forms of I/R injury by attenuating endothelial cell activation/inflammatory response, chemotaxis of inflammatory cells, rolling and adhesion of inflammatory cells to the endothelium, transendothelial migration, adhesion to the parenchymal cells and activation, and interrelated oxidative/nitrosative stress/ inflammatory response (Figure 1).

CB₂ receptor is also detectable in human coronary artery endothelial cells by western blotting, reverse transcription-PCR, real-time PCR and immunofluorescence staining, where its activation by JWH133 or HU-308 attenuates TNFα-induced nuclear factor-κB and RhoA activation, upregulation of adhesion molecules ICAM-1 and vascular cell adhesion molecule-1, increased expression of monocyte chemoattractant protein, enhanced transendothelial migration of monocytes and augmented monocyte-endothelial adhesion (Rajesh et al., 2007a). CB2 agonists also decreased the TNF-α- and/or endotoxin-induced ICAM-1 and vascular cell adhesion molecule-1 expression in isolated aortas and the adhesion of monocytes to aortic vascular endothelium (Rajesh *et al.*, 2007a). Since the above-mentioned TNF- α - and endotoxin-induced phenotypic changes are critical in the initiation and progression of atherosclerosis, these findings suggest that targeting CB₂ receptors on endothelial cells may explain, at least in part, the previously observed beneficial effects of THC in a mouse model of atherosclerosis (Steffens et al., 2005).

Endocannabinoids in I/R: sources, triggers and roles

Previous pioneering studies hypothesized that circulating activated macrophages and platelets are the pivotal sources of endocannabinoids during haemorrhagic shock (Wagner *et al.*, 1997), endotoxemia (Varga *et al.*, 1998), myocardial infarction (Wagner *et al.*, 2001) or liver cirrhosis (Batkai *et al.*, 2001) both in experimental animals and in humans. When isolated and injected into normal rats, these activated cells elicited SR141716-sensitive hypotension, also pointing towards the involvement of CB_1 receptors in many of these conditions (Wagner *et al.*, 1997; Varga *et al.*, 1998; Batkai

et al., 2001; Maccarrone *et al.*, 2001; Ros *et al.*, 2002; Liu *et al.*, 2003).

Several studies have reported increased endocannabinoid levels following cerebral (Schmid et al., 1995; Panikashvili et al., 2001; Schabitz et al., 2002; Berger et al., 2004; Muthian et al., 2004) and hepatic and myocardial I/R injury (Wagner et al., 2001; Kurabayashi et al., 2005; Batkai et al., 2007b); however, the role of endocannabinoids and their sources in I/R injury remains to be a very controversial issue requiring further clarification. Interestingly, a recent study using a rat model of high intraocular pressure-induced retinal I/R found enhanced fatty acid amide hydrolase (FAAH) activity and downregulation of CB1 and transient receptor potential vanilloid-1 (TRPV₁) receptors following I/R (Nucci et al., 2007). The I/R-induced cell death was attenuated either by the FAAH inhibitor URB597 or by the AEA stable analogue methanandamide (MetAEA), suggesting that endogenous AEA tone may play a protective role against injury (Nucci et al., 2007).

In a recent study, we attempted to identify cellular sources and triggers of endocannabinoid production using a mouse model of in vivo hepatic I/R (Batkai et al., 2007b). We found that I/R, but not ischaemia alone, triggered several-fold increases in the hepatic levels of the endocannabinoids AEA and 2-AG, which originated from hepatocytes, Kupffer and endothelial cells. Furthermore, these increases were positively correlated with the degree of tissue damage and serum TNF- α , MIP- 1α and MIP-2 levels. Consistently, brief exposure of primary hepatocytes to various oxidants (H₂O₂, peroxynitrite) or inflammatory stimuli (TNF-α, endotoxin) (Pacher et al., 2006b, 2007) triggered marked increases in cellular endocannabinoid levels. Therefore, the important conclusions of this study are that not only inflammatory stimuli (for example, endotoxin) but also oxidative/nitrosative stress can modulate endocannabinoid levels in hepatocytes, and most likely in most other cell types too. The latter is also supported by recent findings demonstrating that the commonly used chemotherapeutic agent doxorubicin, which is known to mediate its cardiotoxicity by triggering oxidative/ nitrosative stress (Pacher et al., 2003), increased endocannabinoid levels both in the myocardium in vivo and in cardiomyocytes in vitro (Mukhopadhyay et al., 2007). Similarly, up to sixfold increase in endocannabinoid AEA and/or 2-AG levels was observed in the hearts and livers of cirrhotic rats (notable cirrhotic cardiomyopathy is not associated with inflammatory cell infiltration of the myocardium; Batkai et al., 2007a). Therefore, parenchymal cells may also represent a very significant source of endocannabinoids produced in various pathological conditions associated with increased inflammation and/or oxidative tissue injury, in addition to the previously reported activated macrophages (Pacher et al., 2006a). The evidence on changes and possible regulation of endocannabinoid levels in various diseases was recently a subject of excellent overviews (Pertwee, 2005b; Di Marzo and Petrosino, 2007).

Our findings also imply that I/R-induced activation of hepatic endocannabinoids may limit hepatic injury by modulating the expression of adhesion molecules and the infiltration and activation of inflammatory cells by CB₂-dependent/-independent mechanisms, which is also

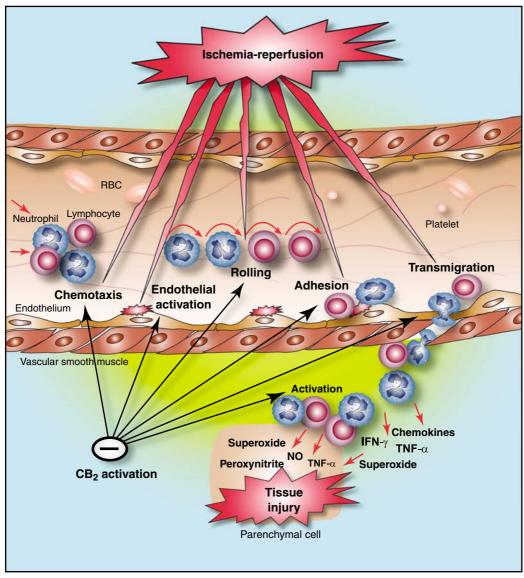


Figure 1 Mechanisms of CB_2 receptor-dependent protection in ischaemia/reperfusion (I/R). CB_2 receptor agonists may protect against I/R injury by attenuating endothelial cell activation/inflammatory response, chemotaxis of inflammatory cells, rolling and adhesion of inflammatory cells to the endothelium, transendothelial migration, adhesion to the parenchymal cells and activation, and interrelated oxidative/nitrosative stress/inflammatory response.

consistent with the emerging role of CB₂ receptors in regulating microglial cell function and neuroinflammation (Walter and Stella, 2004; Maresz *et al.*, 2005). Both mononuclear and polymorphonuclear leukocytes are known to express CB₂ receptors (Klein, 2005; Pacher *et al.*, 2006b), which could be activated on these infiltrating cells through a paracrine mechanism by endocannabinoids generated in and released from the various cell types in the liver. It is noteworthy that the endocannabinoid AEA can promote stellate cell and hepatocyte apoptosis *in vitro* by a mechanism not related to CB receptors (Siegmund *et al.*, 2005, 2006).

Conclusions, future directions

There is a marked increase of endocannabinoid production in various forms of I/R (myocardial, cerebral, hepatic and

circulatory shock), which correlate with the degree of tissue injury and inflammation, and may originate from virtually any cell type involved (Figure 2). Both CB₁ agonists and antagonists may exert various neuroprotective effects against cerebral I/R, the specificity of which should further be studied by using knockout mice. Direct measurements should also confirm the increase of target tissue endocannabinoid levels following preconditioning (most recent evidence is based on assumptions of studies using pharmacological ligands), and experiments using knockout mice should determine the involvement of CB₁/CB₂ receptors. The latter is particularly important, since large body of evidence supporting the idea that CBs may mediate responses via interaction with other sites that probably represent novel CB receptor subtypes such as the putative endothelial CB receptor and GPR55 (Begg et al., 2005; Mackie and Stella, 2006; Hiley and Kaup, 2007; Johns et al., 2007),

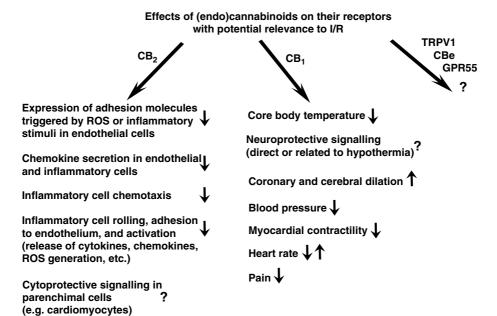


Figure 2 Triggers and sources of endocannabinoids during I/R, and effects mediated via their receptors with potential relevance to pathophysiology. I/R, ischaemia/reperfusion.

and that endocannabinoids may also behave as agonists of $TRPV_1$ receptors under certain conditions and are substrates for COX leading the generation of various biologically active metabolites (Pacher *et al.*, 2005b).

Accumulating recent evidence suggests that selective CB₂ agonists may protect against myocardial, cerebral and hepatic I/R injuries by decreasing the endothelial cell activation/inflammatory response, the expression of adhesion molecules, inflammatory cytokines/chemokines levels, recruitment, adhesion and activation of inflammatory cells, and interrelated oxidative/nitrosative stress. There is considerable interest in the development of selective CB₂ receptor agonists, which are devoid of psychoactive properties of CB₁ agonists, for various inflammatory disorders. Further studies should also establish the therapeutic window of protection during the reperfusion phase with the currently available CB2 receptor agonists, and new compounds should also be designed with better in vivo bioavailability, to devise clinically relevant treatment strategies against various forms of I/R. Nevertheless, the recently observed beneficial effects of CB₂ receptor agonists in hepatic and other forms of I/R, coupled with the absence of psychoactive properties, and antifibrotic effects of CB_2 receptor in the liver suggest that this approach may represent a novel promising strategy against various forms of I/R injury and other inflammatory disorders.

Acknowledgements

This work was supported by the Intramural Research Program of NIH/NIAAA (PP).

Conflict of interest

The authors state no conflict of interest.

References

Ashton JC, Rahman RM, Nair SM, Sutherland BA, Glass M, Appleton I (2007). Cerebral hypoxia-ischemia and middle cerebral artery

- occlusion induce expression of the cannabinoid CB2 receptor in the brain. *Neurosci Lett* **412**: 114–117.
- Batkai S, Jarai Z, Wagner JA, Goparaju SK, Varga K, Liu J et al. (2001). Endocannabinoids acting at vascular CB1 receptors mediate the vasodilated state in advanced liver cirrhosis. Nat Med 7: 827–832.
- Batkai S, Mukhopadhyay P, Harvey-White J, Kechrid R, Pacher P, Kunos G (2007a). Endocannabinoids acting at CB1 receptors mediate the cardiac contractile dysfunction *in vivo* in cirrhotic rats. *Am J Physiol Heart Circ Physiol* **293**: H1689–H1695.
- Batkai S, Osei-Hyiaman D, Pan H, El-Assal O, Rajesh M, Mukhopadhyay P *et al.* (2007b). Cannabinoid-2 receptor mediates protection against hepatic ischemia/reperfusion injury. *FASEB J* 21: 1788–1800.
- Batkai S, Pacher P, Jarai Z, Wagner JA, Kunos G (2004a). Cannabinoid antagonist SR-141716 inhibits endotoxic hypotension by a cardiac mechanism not involving CB1 or CB2 receptors. *Am J Physiol Heart Circ Physiol* **287**: H595–H600.
- Batkai S, Pacher P, Osei-Hyiaman D, Radaeva S, Liu J, Harvey-White J *et al.* (2004b). Endocannabinoids acting at cannabinoid-1 receptors regulate cardiovascular function in hypertension. *Circulation* 110: 1996–2002.
- Begg M, Pacher P, Batkai S, Osei-Hyiaman D, Offertaler L, Mo FM *et al.* (2005). Evidence for novel cannabinoid receptors. *Pharmacol Ther* **106**: 133–145.
- Berger C, Schmid PC, Schabitz WR, Wolf M, Schwab S, Schmid HH (2004). Massive accumulation of *N*-acylethanolamines after stroke. Cell signalling in acute cerebral ischemia? *J Neurochem* 88: 1159–1167.
- Blazquez C, Casanova ML, Planas A, Del Pulgar TG, Villanueva C, Fernandez-Acenero MJ *et al.* (2003). Inhibition of tumor angiogenesis by cannabinoids. *FASEB J* 17: 529–531.
- Bolli R (2007). Preconditioning: a paradigm shift in the biology of myocardial ischemia. *Am J Physiol Heart Circ Physiol* **292**: H19–H27.
- Bouchard JF, Lepicier P, Lamontagne D (2003). Contribution of endocannabinoids in the endothelial protection afforded by ischemic preconditioning in the isolated rat heart. *Life Sci* **72**: 1859–1870.
- Cainazzo MM, Ferrazza G, Mioni C, Bazzani C, Bertolini A, Guarini S (2002). Cannabinoid CB(1) receptor blockade enhances the protective effect of melanocortins in hemorrhagic shock in the rat. *Eur J Pharmacol* **441**: 91–97.
- Carlos TM, Harlan JM (1994). Leukocyte–endothelial adhesion molecules. Blood 84: 2068–2101.
- Croxford JL (2003). Therapeutic potential of cannabinoids in CNS disease. *CNS Drugs* 17: 179–202.
- Di Filippo C, Rossi F, Rossi S, D'Amico M (2004). Cannabinoid CB2 receptor activation reduces mouse myocardial ischemia–reperfusion injury: involvement of cytokine/chemokines and PMN. *J Leukoc Biol* 75: 453–459.
- Di Marzo V, Bifulco M, De Petrocellis L (2004). The endocannabinoid system and its therapeutic exploitation. *Nat Rev Drug Discov* 3: 771–784.
- Di Marzo V, Petrosino S (2007). Endocannabinoids and the regulation of their levels in health and disease. *Curr Opin Lipid* **18**: 129–140.
- Engeli S, Bohnke J, Feldpausch M, Gorzelniak K, Janke J, Batkai S *et al.* (2005). Activation of the peripheral endocannabinoid system in human obesity. *Diabetes* **54**: 2838–2843.
- Evgenov OV, Liaudet L (2005). Role of nitrosative stress and activation of poly(ADP-ribose) polymerase-1 in cardiovascular failure associated with septic and hemorrhagic shock. *Curr Vasc Pharmacol* 3: 293–299.
- Ferdinandy P, Schulz R (2003). Nitric oxide, superoxide, and peroxynitrite in myocardial ischaemia–reperfusion injury and preconditioning. *Br J Pharmacol* **138**: 532–543.
- Freund TF, Katona I, Piomelli D (2003). Role of endogenous cannabinoids in synaptic signalling. *Physiol Rev* 83: 1017–1066.
- Gaskari SA, Liu H, Moezi L, Li Y, Baik SK, Lee SS (2005). Role of endocannabinoids in the pathogenesis of cirrhotic cardiomyopathy in bile duct-ligated rats. *Br J Pharmacol* **146**: 315–323.
- Gebremedhin D, Lange AR, Campbell WB, Hillard CJ, Harder DR (1999). Cannabinoid CB1 receptor of cat cerebral arterial muscle functions to inhibit L-type Ca2+ channel current. *Am J Physiol* **276**: H2085–H2093.

- Golech SA, McCarron RM, Chen Y, Bembry J, Lenz F, Mechoulam R et al. (2004). Human brain endothelium: coexpression and function of vanilloid and endocannabinoid receptors. Brain Res Mol Brain Res 132: 87–92.
- Griendling KK, Sorescu D, Ushio-Fukai M (2000). NAD(P)H oxidase: role in cardiovascular biology and disease. Circ Res 86: 494–501.
- Hampson AJ, Grimaldi M, Lolic M, Wink D, Rosenthal R, Axelrod J (2000). Neuroprotective antioxidants from marijuana. Ann NY Acad Sci 899: 274–282.
- Hausenloy DJ, Yellon DM (2007). Preconditioning and postconditioning: united at reperfusion. *Pharmacol Ther* 116: 173–191.
- Hayakawa K, Mishima K, Abe K, Hasebe N, Takamatsu F, Yasuda H *et al.* (2004). Cannabidiol prevents infarction via the non-CB1 cannabinoid receptor mechanism. *Neuroreport* **15**: 2381–2385.
- Hiley CR, Ford WR (2004). Cannabinoid pharmacology in the cardiovascular system: potential protective mechanisms through lipid signalling. *Biol Rev Camb Philos Soc* **79**: 187–205.
- Hiley CR, Kaup SS (2007). GPR55 and the vascular receptors for cannabinoids. *Br J Pharmacol* **152**: 559–561.
- Hotchkiss RS, Karl IE (2003). The pathophysiology and treatment of sepsis. N Engl J Med 348: 138–150.
- Howlett AC, Barth F, Bonner TI, Cabral G, Casellas P, Devane WA et al. (2002). International union of pharmacology. XXVII. Classification of cannabinoid receptors. Pharmacol Rev 54: 161–202.
- Jaeschke H (1997). Cellular adhesion molecules: regulation and functional significance in the pathogenesis of liver diseases. Am J Physiol 273: G602–G611.
- Jaeschke H (2006). Mechanisms of liver injury. II. Mechanisms of neutrophil-induced liver cell injury during hepatic ischemiareperfusion and other acute inflammatory conditions. Am J Physiol 290: G1083–G1088.
- Jaeschke H, Smith CW, Clemens MG, Ganey PE, Roth RA (1996). Mechanisms of inflammatory liver injury: adhesion molecules and cytotoxicity of neutrophils. *Toxicol Applied Pharmacol* 139: 213–226.
- Johns DG, Behm DJ, Walker DJ, Ao Z, Shapland EM, Daniels DA *et al.* (2007). The novel endocannabinoid receptor GPR55 is activated by atypical cannabinoids but does not mediate their vasodilator effects. *Br J Pharmacol* **152**: 825–831.
- Joyeux M, Arnaud C, Godin-Ribuot D, Demenge P, Lamontagne D, Ribuot C (2002). Endocannabinoids are implicated in the infarct size-reducing effect conferred by heat stress preconditioning in isolated rat hearts. *Cardiovas Res* 55: 619–625.
- Julien B, Grenard P, Teixeira-Clerc F, Van Nhieu JT, Li L, Karsak M et al. (2005). Antifibrogenic role of the cannabinoid receptor CB2 in the liver. Gastroenterology 128: 742–755.
- Kadoi Y, Goto F (2006). Effects of AM281, a cannabinoid antagonist, on circulatory deterioration and cytokine production in an endotoxin shock model: comparison with norepinephrine. *J Anesth* 20: 284–289.
- Kadoi Y, Hinohara H, Kunimoto F, Saito S, Goto F (2005). Cannabinoid antagonist AM 281 reduces mortality rate and neurologic dysfunction after cecal ligation and puncture in rats. Crit Care Med 33: 2629–2636.
- Klein TW (2005). Cannabinoid-based drugs as anti-inflammatory therapeutics. *Nat Rev Immunol* 5: 400–411.
- Klijn CJ, Hankey GJ (2003). Management of acute ischaemic stroke: new guidelines from the American Stroke Association and European Stroke Initiative. *Lancet Neurol* 2: 698–701.
- Krylatov AV, Bernatskaia NA, Maslov LN, Pertwee RG, Mechoulam R, Stefano GB *et al.* (2002a). Increase of the heart arrhythmogenic resistance and decrease of the myocardial necrosis zone during activation of cannabinoid receptors. *Ross Fiziol Zh Im I M Sechenova* 88: 560–567.
- Krylatov AV, Ugdyzhekova DS, Bernatskaya NA, Maslov LN, Mekhoulam R, Pertwee RG et al. (2001). Activation of type II cannabinoid receptors improves myocardial tolerance to arrhythmogenic effects of coronary occlusion and reperfusion. Bull Exp Biol and Med 131: 523–525.
- Krylatov AV, Uzhachenko RV, Maslov LN, Ugdyzhekova DS, Bernatskaia NA, Pertwee R *et al.* (2002b). Anandamide and R-(+)-methanandamide prevent development of ischemic and reperfusion arrhythmia in rats by stimulation of CB2-receptors. *Eksp Klin Farmakol* 65: 6–9.

- Kurabayashi M, Takeyoshi I, Yoshinari D, Matsumoto K, Maruyama I, Morishita Y (2005). 2-Arachidonoylglycerol increases in ischemiareperfusion injury of the rat liver. J Invest Surg 18: 25–31.
- Lagneux *C*, Lamontagne D (2001). Involvement of cannabinoids in the cardioprotection induced by lipopolysaccharide. *Br J Pharmacol* **132**: 793–796.
- Lamontagne D, Lepicier P, Lagneux C, Bouchard JF (2006). The endogenous cardiac cannabinoid system: a new protective mechanism against myocardial ischemia. Arch Mal Coeur Vaiss 99: 242–246.
- Lees KR, Davalos A, Davis SM, Diener HC, Grotta J, Lyden P *et al.* (2006). Additional outcomes and subgroup analyses of NXY-059 for acute ischemic stroke in the SAINT I trial. *Stroke* **37**: 2970–2978.
- Leker RR, Gai N, Mechoulam R, Ovadia H (2003). Drug-induced hypothermia reduces ischemic damage: effects of the cannabinoid HU-210. Stroke 34: 2000–2006.
- Lepicier P, Bibeau-Poirier A, Lagneux C, Servant MJ, Lamontagne D (2006). Signalling pathways involved in the cardioprotective effects of cannabinoids. *J Pharmacol Sci* 102: 155–166.
- Lepicier P, Bouchard JF, Lagneux C, Lamontagne D (2003). Endocannabinoids protect the rat isolated heart against ischaemia. *Br J Pharmacol* **139**: 805–815.
- Liu J, Batkai S, Pacher P, Harvey-White J, Wagner JA, Cravatt BF *et al.* (2003). Lipopolysaccharide induces anandamide synthesis in macrophages via CD14/MAPK/phosphoinositide 3-kinase/NF-kappaB independently of platelet-activating factor. *J Biol Chem* **278**: 45034–45039.
- Liu J, Gao B, Mirshahi F, Sanyal AJ, Khanolkar AD, Makriyannis A *et al.* (2000). Functional CB1 cannabinoid receptors in human vascular endothelial cells. *Biochem J* **346** (Part 3): 835–840.
- Liu J, Wang L, Harvey-White J, Osei-Hyiaman D, Razdan R, Gong Q et al. (2006). A biosynthetic pathway for anandamide. Proc Natl Acad Sci USA 103: 13345–13350.
- Louw DF, Yang FW, Sutherland GR (2000). The effect of delta-9-tetrahydrocannabinol on forebrain ischemia in rat. *Brain Res* **857**: 183–187.
- Lucchesi BR (1990). Modulation of leukocyte-mediated myocardial reperfusion injury. Annu Rev Physiol 52: 561–576.
- Maccarrone M, De Petrocellis L, Bari M, Fezza F, Salvati S, Di Marzo V *et al.* (2001). Lipopolysaccharide downregulates fatty acid amide hydrolase expression and increases anandamide levels in human peripheral lymphocytes. *Arc Biochem Biophys* **393**: 321–328.
- Mackie K (2006). Cannabinoid receptors as therapeutic targets. Ann Rev Pharmacol Toxicol 46: 101–122.
- Mackie K, Stella N (2006). Cannabinoid receptors and endocannabinoids: evidence for new players. AAPS J 8: E298–E306.
- Mallat A, Teixeira-Clerc F, Deveaux V, Lotersztajn S (2007). Cannabinoid receptors as new targets of antifibrosing strategies during chronic liver diseases. *Exp Opin Ther Targets* 11: 403–409.
- Maresz K, Carrier EJ, Ponomarev ED, Hillard CJ, Dittel BN (2005). Modulation of the cannabinoid CB2 receptor in microglial cells in response to inflammatory stimuli. *J Neurochem* 95: 437–445.
- Matsuda K, Mikami Y, Takeda K, Fukuyama S, Egawa S, Sunamura M *et al.* (2005). The cannabinoid 1 receptor antagonist, AM251, prolongs the survival of rats with severe acute pancreatitis. *Tohoku J Exp Med* **207**: 99–107.
- Matsuda LA, Lolait SJ, Brownstein MJ, Young AC, Bonner TI (1990). Structure of a cannabinoid receptor and functional expression of the cloned cDNA. *Nature* **346**: 561–564.
- Mauler F, Mittendorf J, Horvath E, De Vry J (2002). Characterization of the diarylether sulfonylester (–)-(R)-3-(2-hydroxymethylindanyl-4-oxy)phenyl-4,4,4-trifluoro-1-sulfonate (BAY 38-7271) as a potent cannabinoid receptor agonist with neuroprotective properties. *J Pharml Exp Ther* 302: 359–368.
- Mechoulam R, Panikashvili D, Shohami E (2002a). Cannabinoids and brain injury: therapeutic implications. *Trends Mol Med* 8: 58–61.
- Mechoulam R, Spatz M, Shohami E (2002b). Endocannabinoids and neuroprotection. *Sci STKE* **2002**: RE5.
- Mendizabal VE, Adler-Graschinsky E (2007). Cannabinoids as therapeutic agents in cardiovascular disease: a tale of passions and illusions. *Br J Pharmacol* **151**: 427–440.
- Mestre L, Correa F, Docagne F, Clemente D, Guaza C (2006). The synthetic cannabinoid WIN 55,212-2 increases COX-2 expression

- and PGE2 release in murine brain-derived endothelial cells following Theiler's virus infection. *Biochem Pharmacol* **72**: 869–880.
- Moezi L, Gaskari SA, Liu H, Baik SK, Dehpour AR, Lee SS (2006). Anandamide mediates hyperdynamic circulation in cirrhotic rats via CB(1) and VR(1) receptors. *Br J Pharmacol* **149**: 898–908.
- Mukhopadhyay P, Batkai S, Rajesh M, Czifra N, Harvey-White J, Hasko G *et al.* (2007). Pharmacological inhibition of cannabinoid receptor-1 protects against doxorubicin-induced cardiotoxicity. *J Am Coll Cardiol* **50**: 528–536.
- Munro S, Thomas KL, Abu-Shaar M (1993). Molecular characterization of a peripheral receptor for cannabinoids. *Nature* **365**: 61–65.
- Murry CE, Jennings RB, Reimer KA (1986). Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation* **74**: 1124–1136.
- Muthian S, Rademacher DJ, Roelke CT, Gross GJ, Hillard CJ (2004). Anandamide content is increased and CB1 cannabinoid receptor blockade is protective during transient, focal cerebral ischemia. *Neuroscience* **129**: 743–750.
- Nagayama T, Sinor AD, Simon RP, Chen J, Graham SH, Jin K et al. (1999). Cannabinoids and neuroprotection in global and focal cerebral ischemia and in neuronal cultures. J Neurosci 19: 2987–2995
- Nucci C, Gasperi V, Tartaglione R, Cerulli A, Terrinoni A, Bari M *et al.* (2007). Involvement of the endocannabinoid system in retinal damage after high intraocular pressure-induced ischemia in rats. *Invest Ophthalmol Vis Sci* **48**: 2997–3004.
- Ohkohchi N, Shibuya H, Tsukamoto S, Sakurada M, Oikawa K, Terashima T *et al.* (1999). Kupffer's cells modulate neutrophile activity by superoxide anion and tumor necrosis factor-delta in reperfusion injury of liver transplantation-mechanisms of radical generation and reperfusion injury after cold ischemia. *Transp Proc* 31: 1055–1058.
- Osei-Hyiaman D, DePetrillo M, Pacher P, Liu J, Radaeva S, Batkai S *et al.* (2005). Endocannabinoid activation at hepatic CB1 receptors stimulates fatty acid synthesis and contributes to diet-induced obesity. *J Clin Invest* 115: 1298–1305.
- Pacher P, Batkai S, Kunos G (2005a). Blood pressure regulation by endocannabinoids and their receptors. *Neuropharmacology* 48: 1130–1138.
- Pacher P, Batkai S, Kunos G (2005b). Cardiovascular pharmacology of cannabinoids. *Handb Exp Pharmacol* **168**: 599–625.
- Pacher P, Batkai S, Kunos G (2005c). Cirrhotic cardiomyopathy: an endocannabinoid connection? *Br J Pharmacol* **146**: 313–314.
- Pacher P, Batkai S, Kunos G (2006a). The endocannabinoid system as an emerging target of pharmacotherapy. *Pharmacol Rev* 58: 389 462
- Pacher P, Batkai S, Osei-Hyiaman D, Offertaler L, Liu J, Harvey-White J *et al.* (2005d). Hemodynamic profile, responsiveness to anandamide, and baroreflex sensitivity of mice lacking fatty acid amide hydrolase. *Am J Physiol Heart Circ Physiol* **289**: H533–H541.
- Pacher P, Beckman JS, Liaudet L (2007). Nitric oxide and peroxynitrite in health and disease. *Physiol Rev* 87: 315–424.
- Pacher P, Liaudet L, Bai P, Mabley JG, Kaminski PM, Virag L *et al.* (2003). Potent metalloporphyrin peroxynitrite decomposition catalyst protects against the development of doxorubicin-induced cardiac dysfunction. *Circulation* **107**: 896–904.
- Pacher P, Nivorozhkin A, Szabo C (2006b). Therapeutic effects of xanthine oxidase inhibitors: renaissance half a century after the discovery of allopurinol. *Pharmacol Rev* 58: 87–114.
- Pacher P, Schulz R, Liaudet L, Szabo C (2005e). Nitrosative stress and pharmacological modulation of heart failure. *Trends Pharmacol Sci* **26**: 302–310.
- Panikashvili D, Mechoulam R, Beni SM, Alexandrovich A, Shohami E (2005). CB1 cannabinoid receptors are involved in neuroprotection via NF-kappa B inhibition. J Cereb Blood Flow Metab 25: 477–484.
- Panikashvili D, Shein NA, Mechoulam R, Trembovler V, Kohen R, Alexandrovich A et al. (2006). The endocannabinoid 2-AG protects the blood–brain barrier after closed head injury and inhibits mRNA expression of proinflammatory cytokines. Neurobiol Dis 22: 257–264.
- Panikashvili D, Simeonidou C, Ben-Shabat S, Hanus L, Breuer A, Mechoulam R et al. (2001). An endogenous cannabinoid (2-AG) is neuroprotective after brain injury. Nature 413: 527–531.

- Parmentier-Batteur S, Jin K, Mao XO, Xie L, Greenberg DA (2002). Increased severity of stroke in CB1 cannabinoid receptor knock-out mice. *J Neurosci* 22: 9771–9775.
- Pertwee RG (2005a). Pharmacological actions of cannabinoids. *Handb Exp Pharmacol* **168**: 1–51.
- Pertwee RG (2005b). The therapeutic potential of drugs that target cannabinoid receptors or modulate the tissue levels or actions of endocannabinoids. *AAPS J* 7: E625–E654.
- Pertwee RG (2006). The pharmacology of cannabinoid receptors and their ligands: an overview. *Int J Obes (Lond)* 30 (Suppl 1): S13–S18.
- Piomelli D (2003). The molecular logic of endocannabinoid signalling. *Nat Rev Neurosci* 4: 873–884.
- Podgoreanu MV, Michelotti GA, Sato Y, Smith MP, Lin S, Morris RW *et al.* (2005). Differential cardiac gene expression during cardio-pulmonary bypass: ischemia-independent upregulation of proinflammatory genes. *J Thorac Cardiovasc Surg* **130**: 330–339.
- Rajesh M, Mukhopadhyay P, Batkai S, Hasko G, Liaudet L, Huffman JW *et al.* (2007a). CB2 cannabinoid receptor stimulation attenuates TNF-α-induced human endothelial cell activation, transendothelial migration of monocytes, and monocyte–endothelial adhesion. *Am J Physiol Heart Circ Physiol* **293**: H2210–H2218.
- Rajesh M, Pan H, Mukhopadhyay P, Batkai S, Osei-Hyiaman D, Hasko G et al. (2007b). Cannabinoid-2 receptor agonist HU-308 protects against hepatic ischemia/reperfusion injury by attenuating oxidative stress, inflammatory response and apoptosis. *J Leukoc Biol* 1 December, 82 (6).
- Randall MD, Harris D, Kendall DA, Ralevic V (2002). Cardiovascular effects of cannabinoids. *Pharmacol Ther* 95: 191–202.
- Ros J, Claria J, To-Figueras J, Planaguma A, Cejudo-Martin P, Fernandez-Varo G *et al.* (2002). Endogenous cannabinoids: a new system involved in the homeostasis of arterial pressure in experimental cirrhosis in the rat. *Gastroenterology* **122**: 85–93.
- Sarne Y, Mechoulam R (2005). Cannabinoids: between neuroprotection and neurotoxicity. Curr Drug Targets CNS Neurol Disord 4: 677–684.
- Schabitz WR, Giuffrida A, Berger C, Aschoff A, Schwaninger M, Schwab S et al. (2002). Release of fatty acid amides in a patient with hemispheric stroke: a microdialysis study. Stroke 33: 2112–2114.
- Schmid PC, Krebsbach RJ, Perry SR, Dettmer TM, Maasson JL, Schmid HH (1995). Occurrence and postmortem generation of anandamide and other long-chain *N*-acylethanolamines in mammalian brain. *FEBS Lett* **375**: 117–120.
- Shmist YA, Goncharov I, Eichler M, Shneyvays V, Isaac A, Vogel Z *et al.* (2006). Delta-9-tetrahydrocannabinol protects cardiac cells from hypoxia via CB2 receptor activation and nitric oxide production. *Mol Cell Biochem* **283**: 75–83.
- Siegmund SV, Seki E, Osawa Y, Uchinami H, Cravatt BF, Schwabe RF (2006). Fatty acid amide hydrolase determines anandamideinduced cell death in the liver. J Biol Chem 281: 10431–10438.
- Siegmund SV, Uchinami H, Osawa Y, Brenner DA, Schwabe RF (2005). Anandamide induces necrosis in primary hepatic stellate cells. *Hepatology* **41**: 1085–1095.
- Sinor AD, Irvin SM, Greenberg DA (2000). Endocannabinoids protect cerebral cortical neurons from *in vitro* ischemia in rats. *Neurosci Lett* 278: 157–160.
- Smith SR, Denhardt G, Terminelli C (2001). The anti-inflammatory activities of cannabinoid receptor ligands in mouse peritonitis models. *Eur J Pharmacol* **432**: 107–119.
- Smith SR, Terminelli C, Denhardt G (2000). Effects of cannabinoid receptor agonist and antagonist ligands on production of inflammatory cytokines and anti-inflammatory interleukin-10 in endotoxemic mice. *J Pharmacol Exp Ther* **293**: 136–150.
- Sommer C, Schomacher M, Berger C, Kuhnert K, Muller HD, Schwab S et al. (2006). Neuroprotective cannabinoid receptor antagonist SR141716A prevents downregulation of excitotoxic NMDA receptors in the ischemic penumbra. Acta Neuropathol 112: 277–286.

- Steffens S, Veillard NR, Arnaud C, Pelli G, Burger F, Staub C *et al.* (2005). Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. *Nature* **434**: 782–786.
- Teixeira-Clerc F, Julien B, Grenard P, Tran Van Nhieu J, Deveaux V, Li L *et al.* (2006). CB1 cannabinoid receptor antagonism: a new strategy for the treatment of liver fibrosis. *Nat Med* **12**: 671–676.
- Ugdyzhekova DS, Bernatskaya NA, Stefano JB, Graier VF, Tam SW, Mekhoulam R (2001). Endogenous cannabinoid anandamide increases heart resistance to arrhythmogenic effects of epinephrine: role of CB(1) and CB(2) receptors. *Bull Exp Biol Med* 131: 251–253
- Ugdyzhekova DS, Krylatov AV, Bernatskaya NA, Maslov LN, Mechoulam R, Pertwee RG (2002). Activation of cannabinoid receptors decreases the area of ischemic myocardial necrosis. *Bull Exp Biol Med* 133: 125–126.
- Underdown NJ, Hiley CR, Ford WR (2005). Anandamide reduces infarct size in rat isolated hearts subjected to ischaemia–reperfusion by a novel cannabinoid mechanism. *Br J Pharmacol* **146**: 809–816.
- Ungvari Z, Gupte SA, Recchia FA, Batkai S, Pacher P (2005). Role of oxidative–nitrosative stress and downstream pathways in various forms of cardiomyopathy and heart failure. Curr Vasc Pharmacol 3: 221–229.
- Van Sickle MD, Duncan M, Kingsley PJ, Mouihate A, Urbani P, Mackie K *et al.* (2005). Identification and functional characterization of brainstem cannabinoid CB2 receptors. *Science* **310**: 329–332.
- Varga K, Wagner JA, Bridgen DT, Kunos G (1998). Platelet- and macrophage-derived endogenous cannabinoids are involved in endotoxin-induced hypotension. FASEB J 12: 1035–1044.
- Wagner JA, Abesser M, Harvey-White J, Ertl G (2006). 2-Arachidonylglycerol acting on CB1 cannabinoid receptors mediates delayed cardioprotection induced by nitric oxide in rat isolated hearts. *J Cardiovasc Pharm* 47: 650–655.
- Wagner JA, Hu K, Bauersachs J, Karcher J, Wiesler M, Goparaju SK et al. (2001). Endogenous cannabinoids mediate hypotension after experimental myocardial infarction. J Am Coll Cardiol 38: 2048–2054.
- Wagner JA, Hu K, Karcher J, Bauersachs J, Schafer A, Laser M *et al.* (2003). CB(1) cannabinoid receptor antagonism promotes remodeling and cannabinoid treatment prevents endothelial dysfunction and hypotension in rats with myocardial infarction. *Br J Pharmacol* 138: 1251–1258.
- Wagner JA, Varga K, Ellis EF, Rzigalinski BA, Martin BR, Kunos G (1997). Activation of peripheral CB1 cannabinoid receptors in haemorrhagic shock. *Nature* **390**: 518–521.
- Walter L, Stella N (2004). Cannabinoids and neuroinflammation. Br J Pharmacol 141: 775–785.
- Wiley JL, Beletskaya ID, Ng EW, Dai Z, Crocker PJ, Mahadevan A *et al.* (2002). Resorcinol derivatives: a novel template for the development of cannabinoid CB(1)/CB(2) and CB(2)-selective agonists. *J Pharmacol Exp Ther* **301**: 679–689.
- Yang YY, Lin HC, Huang YT, Lee TY, Hou MC, Wang YW *et al.* (2007). Effect of chronic CB1 cannabinoid receptor antagonism on livers of rats with biliary cirrhosis. *Clin Sci (Lond)* **112**: 533–542.
- Yellon DM, Hausenloy DJ (2005). Realizing the clinical potential of ischemic preconditioning and postconditioning. Nat Clin Pract Cardiovasc Med 2: 568–575.
- Zhang M, Martin BR, Adler MW, Razdan RK, Jallo JI, Tuma RF (2007). Cannabinoid CB(2) receptor activation decreases cerebral infarction in a mouse focal ischemia/reperfusion model. *J Cereb Blood Flow Metab* 27: 1387–1396.
- Zoratti C, Kipmen-Korgun D, Osibow K, Malli R, Graier WF (2003). Anandamide initiates Ca(2+) signalling via CB2 receptor linked to phospholipase C in calf pulmonary endothelial cells. *Br J Pharmacol* **140**: 1351–1362.